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# High-throughput SPR assay for FcγR binding to drug candidates on the new Carterra LSA<sup>XT</sup>

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## About me / About Dragonfly

- Graduated Northeastern with B.S. in Bioengineering in 2022
- Associate Scientist at Dragonfly for 2+ years
- Dragonfly founded in 2016
  - Multiple clinical-stage therapies using TriNKET and cytokine platforms
  - Partners include BMS, Merck, Gilead, AbbVie



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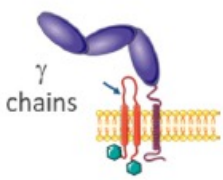

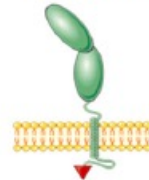



## Outline

- Importance of Fc $\gamma$ R characterization for Fc-containing molecules
- LSA<sup>XT</sup> implementation for Fc $\gamma$ R screening
- Human and cynomolgus assay design and results
- Comparison of LSA<sup>XT</sup> to Biacore 8K
- Conclusions



# Fc gamma receptor (Fc $\gamma$ R) binding is a critical component in antibody mediated immune activity

- Fc $\gamma$ Rs are essential in ADCC, ADCP, and CDC activity.
- Fc $\gamma$ R binding should be characterized for all Fc-containing molecules.

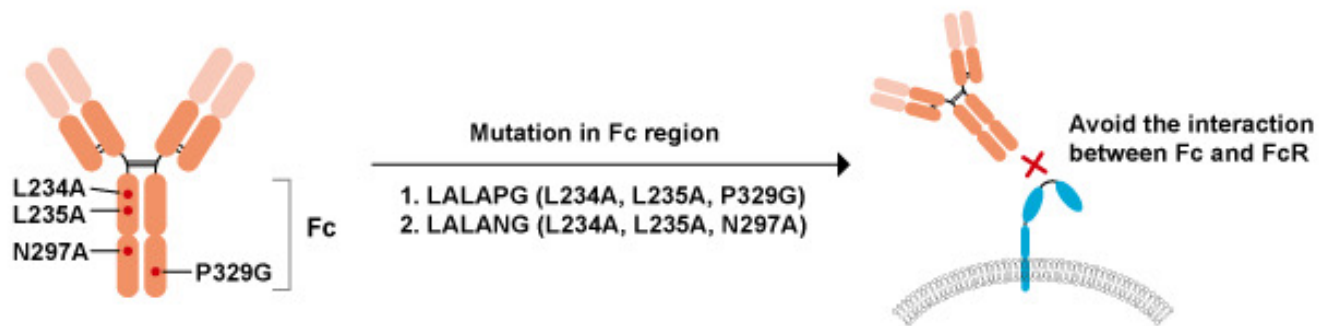
Receptor	Fc $\gamma$ RI		Fc $\gamma$ RII		Fc $\gamma$ RIII	
	Fc $\gamma$ RIa	Fc $\gamma$ RIIa	Fc $\gamma$ RIIb	Fc $\gamma$ RIIc	Fc $\gamma$ RIIIa	Fc $\gamma$ RIIIb
Structure						
Signal	ITAM	ITAM	ITIM	ITAM	ITAM	-
Polymorphism	-	R131H	I232T	Q57X	V158F	NA1, NA2, SH
Affinity for IgG	High (10 <sup>-8</sup> -10 <sup>-9</sup> M)		Low (10 <sup>-6</sup> M)		Low (10 <sup>-6</sup> -10 <sup>-7</sup> M)	



# Fc $\gamma$ R binding analysis is crucial in antibody and Fc-fusion therapeutic development

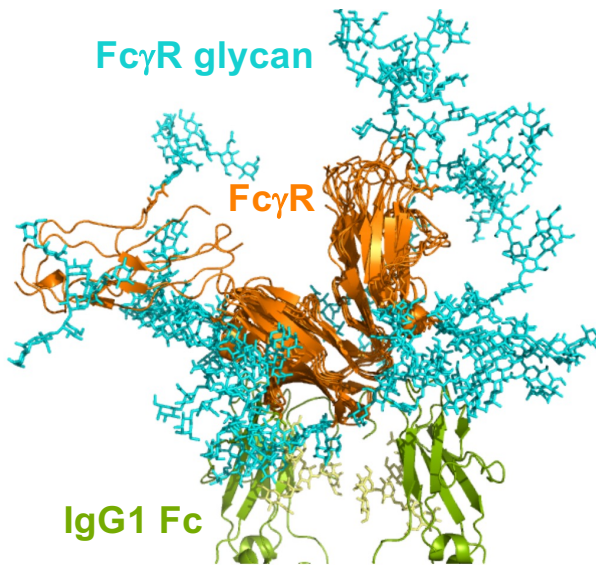
Fc $\gamma$ R binding assessment provides critical information into wild-type or engineered Fc domains. Routine assessments include:

1. Wild-type Fc $\gamma$ R binding determination
2. Cross-reactivity to relevant species Fc $\gamma$ Rs
3. Impact of Fc mutations (Fc $\gamma$ R silencing, enhancing, etc.)





# Mutations augmenting effector function modulate binding affinity for multiple FcγRs



Fc Modification	ID	Effector Function vs. WT		Binding Affinity vs. WT				
		ADCC	ADCP	FcγRIIA <sup>H</sup>	FcγRIIA <sup>R</sup>	FcγRIIB	FcγRIIIA <sup>F</sup>	FcγRIIIA <sup>V</sup>
S298A/E333A/K334A	AAA	↑	n.d.	↓↓	n.d.	↓↓	↑↑	↑
Afucosylation (Potelligent)	Potelligent	↑↑	n.d.	n.d.	n.d.	n.d.	↑↑↑	n.d.
S239D/I332E	DE	↑↑	↑	n.d.	n.d.	↑↑↑	↑↑↑	↑↑↑↑
S239D/A330L/I332E	DLE	↑↑↑	↑	n.d.	n.d.	↑↑↑	↑↑↑	↑↑↑↑
G236A	G236A	-	↑	↑↑	↑↑	↑	-	-
G236A/S239D/I332E	ADE	↑	↑	↑↑	↑↑↑	↑↑↑	↑↑↑	↑↑↑
G236A/A330L/I332E	GAALIE	d.n.s.	n.d.	↑↑	↑↑	↓↓	↑↑	↑↑
G236A/S239D/A330L/I332E	GASDALIE	n.d.	n.d.	↑↑↑	↑↑↑	↑	↑↑↑	↑
F243L/R292P/Y300L/V305I/P396L	LPLIL	↑↑	n.d.	↑↑	↑	↑	↑↑↑	↑↑↑
L235V/F243L/R292P/Y300L/P396L	VLPLL	↑	n.d.	↑	↓↓	↓↓	↑↑	↑

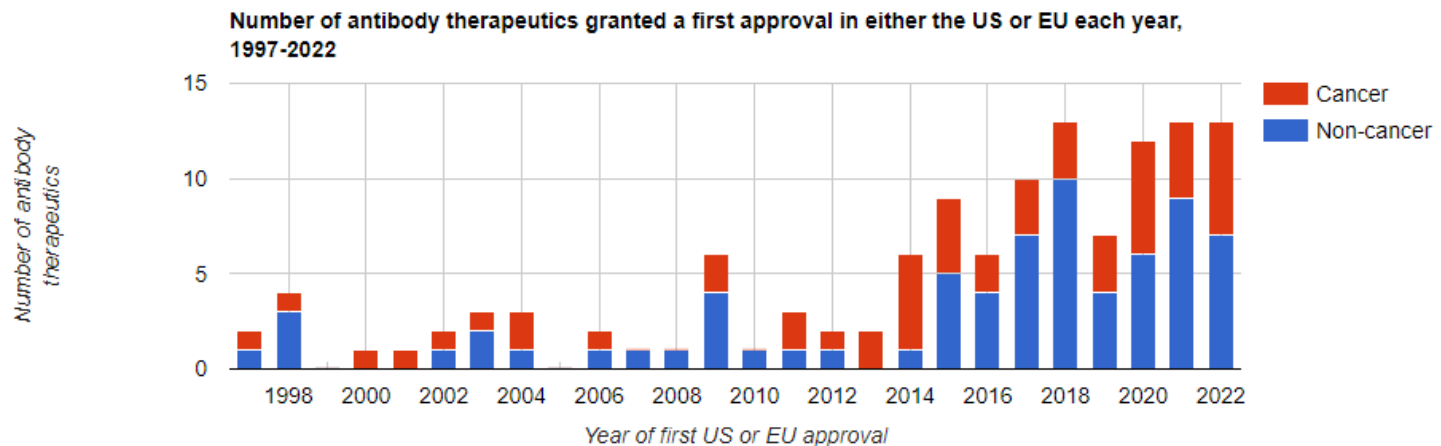
Hayes *et al.* (Journal of Inflammation Research, 2016)

Liu *et al.* (Antibodies, 2020)



## Fc-containing biologics are a substantial and growing class of drugs

- 175+ antibody therapeutics approved or under regulatory review globally
- US market alone valued at \$180 billion (2021)
  - Compound annual growth rate of 14%





## Why consider improving the Fc $\gamma$ R characterization workflow at Dragonfly?

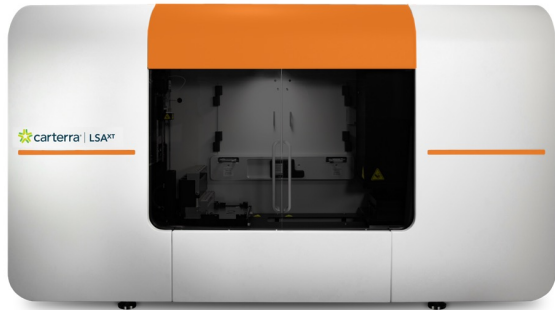
- Fc $\gamma$ R characterization time:
  - Screening human/cyno Fc $\gamma$ R panel on 8K takes one week
- User lab time:
  - Each day a new experiment must be set up
- Protein requirements:
  - Nearly 16mg of each antibody required
- Sensor chips:
  - 6 chips required per antibody





# Could Carterra LSA<sup>XT</sup> help with the Fc $\gamma$ R workflow at Dragonfly?

100x the data | 10% the time | 1% the sample, *plus:*



Enhanced signal-to-noise



Increased signal uniformity



Faster data collection rate

Ability to reduce surface densities

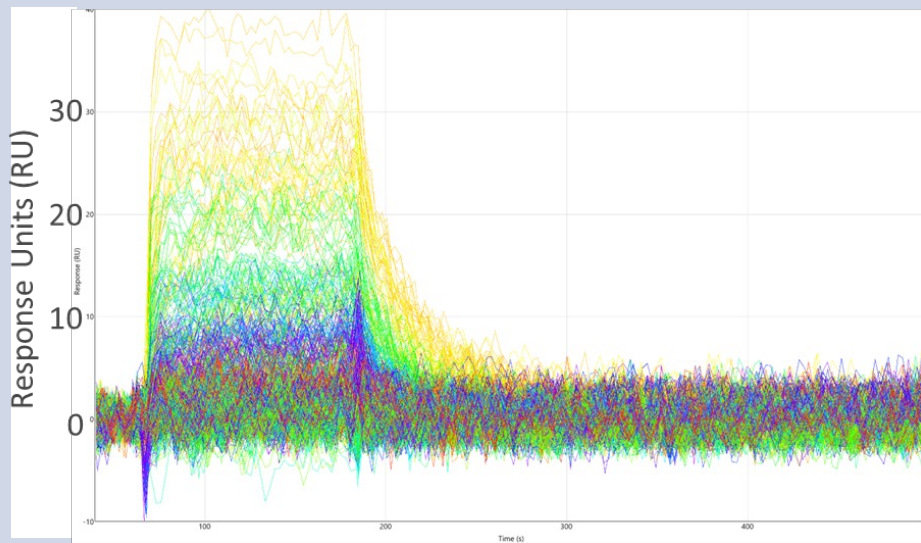
Confidence in replicates

Accurately describe rapid kinetics

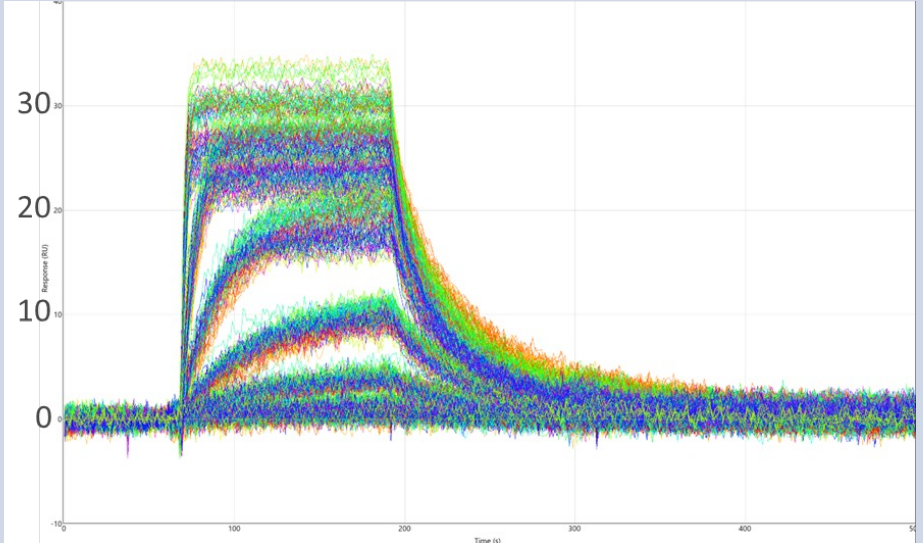


# LSA<sup>XT</sup> demonstrates increased data resolution and reproducibility over LSA

## IL-2 binding to IL2R $\alpha$



LSA  
12 replicates overlaid

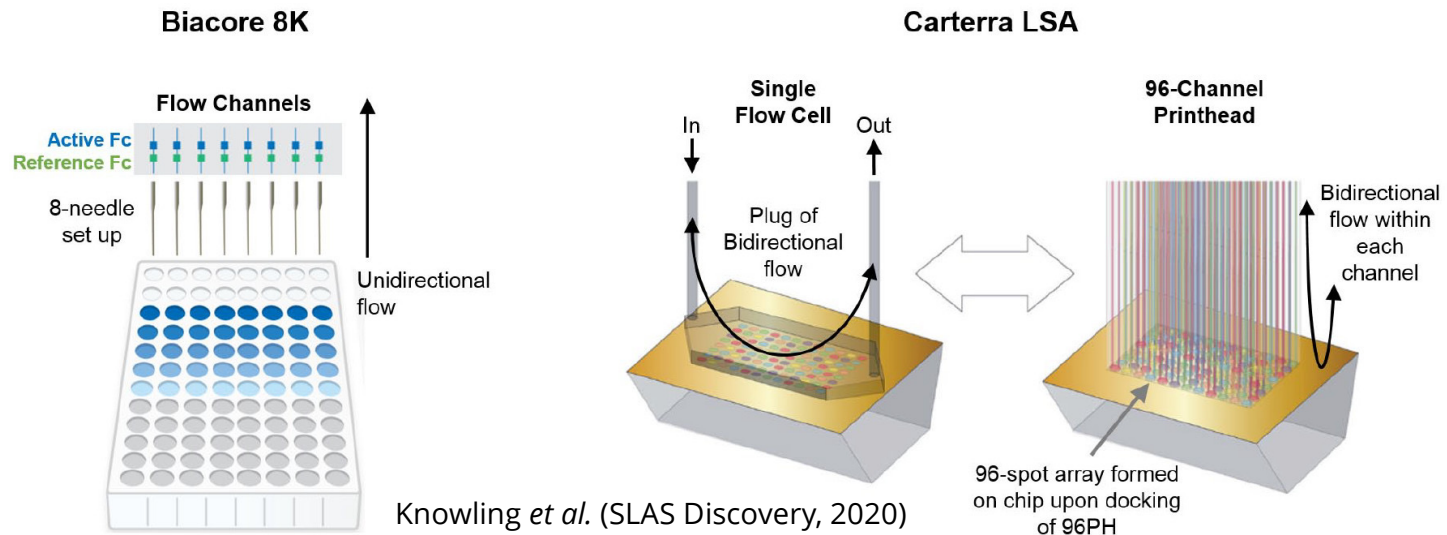


LSA<sup>XT</sup>  
96 replicates overlaid



# Major differences in instrument design allow for higher throughput with Carterra LSA<sup>XT</sup>

- Biacore 8K provides high-resolution kinetics, but flow cell design limits ligand capacity to 8.
- Carterra LSA<sup>XT</sup>'s flow cell design can capture 384 unique ligands with now improved sensitivity.





## Similar assay conditions were established to directly compare between instruments

- Test trastuzumab (IgG1 mAb) binding against 11 Fc $\gamma$ Rs.
- Both instruments ran at 22°C with streptavidin-coated chips.
- All receptors are biotinylated and provided by ACRO Biosystems.



Species	Receptor Tested
Human	CD16a (V & F)
Cynomolgus	CD16
Human	CD16b (NA1 & NA2)
Human	CD32a (H & R)
Human & Cynomolgus	CD32b
Human & Cynomolgus	CD64*

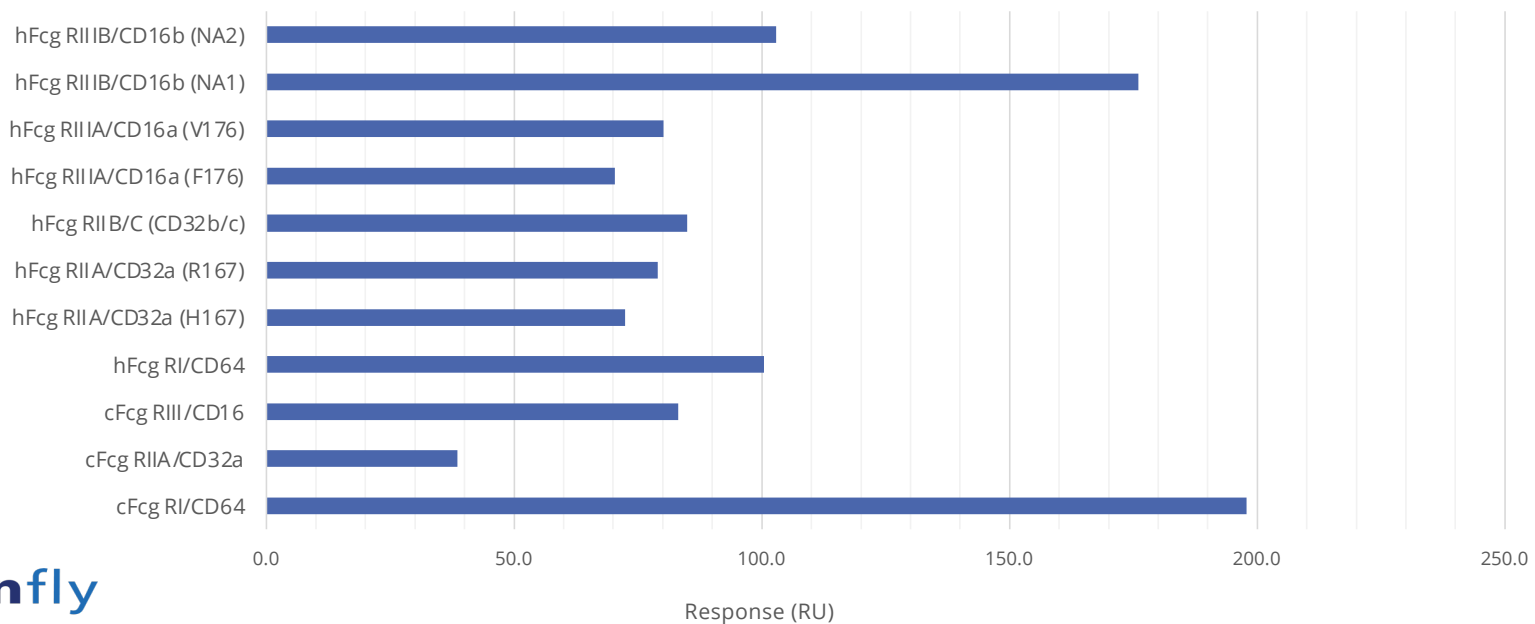
\*Performed on Biacore biotin CAPture chip



# LSA<sup>XT</sup> can capture full FcγR array at a consistent and low surface density

- 11 biotinylated receptors captured on SAHC30M sensor chip
- Standard deviation of capture response replicates  $\leq 15\%$  (n=8)

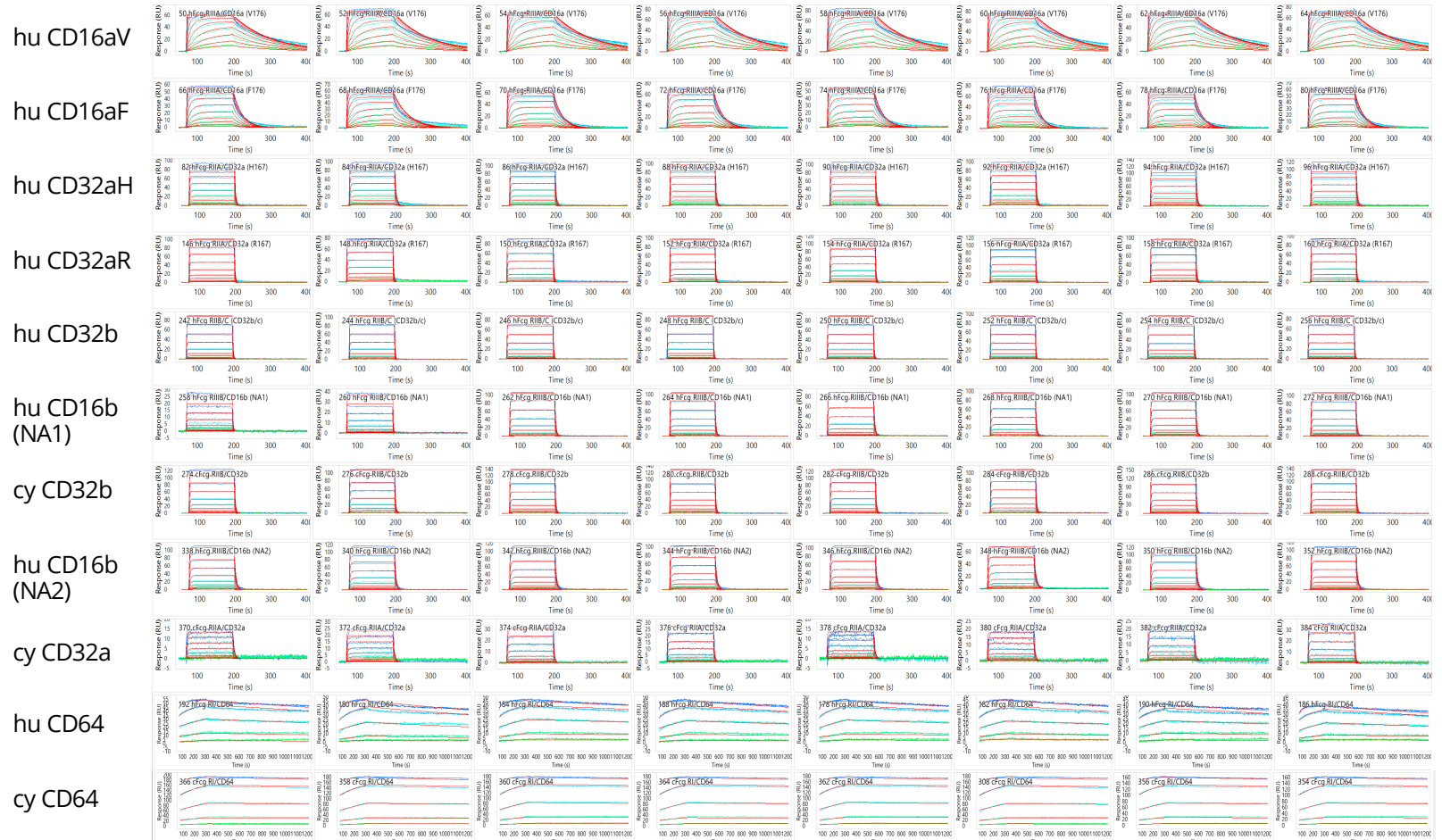
Average capture levels of biotinylated FcγRs





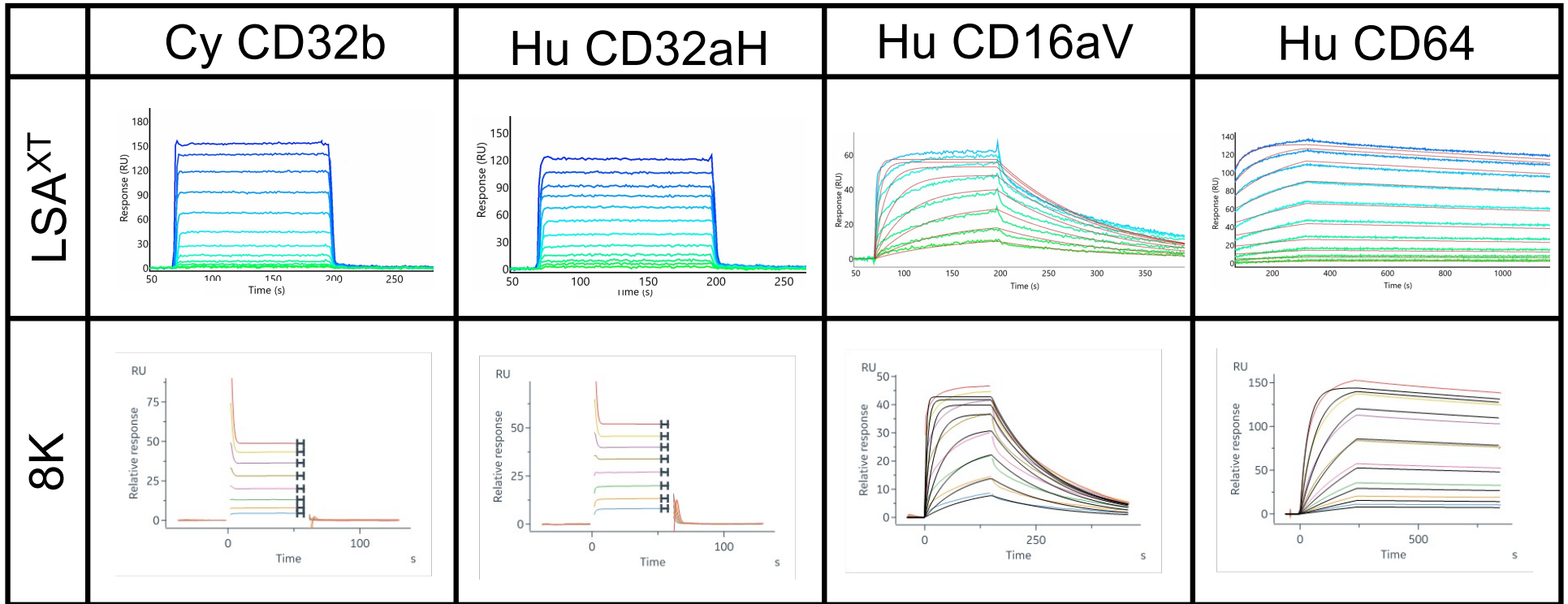
# Fc Gamma Receptor Affinity Characterization on LSA<sup>XT</sup>

11 canonical human and cyno receptors screened against trastuzumab (n=8)



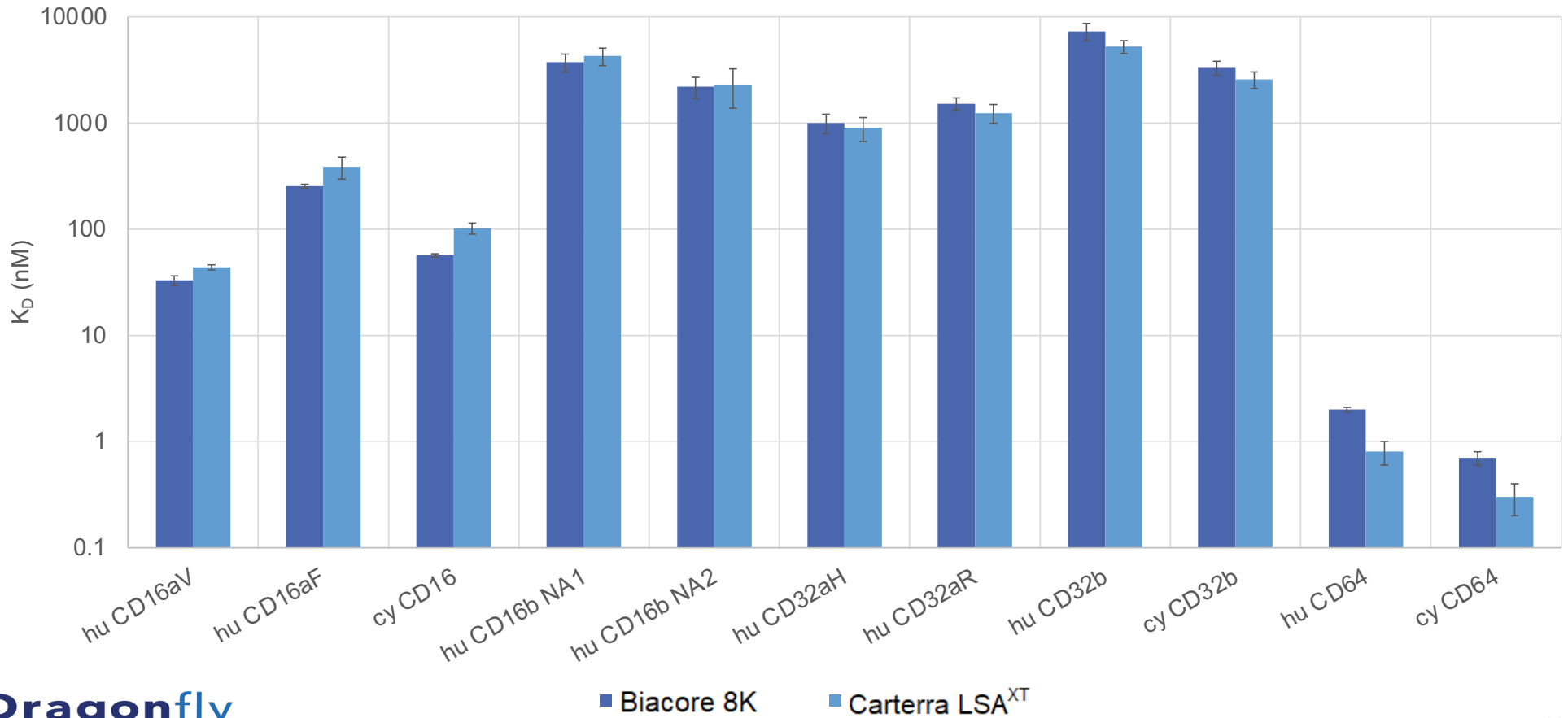


# LSA<sup>XT</sup> and 8K provide high-resolution sensorgrams





# LSA<sup>XT</sup> and 8K produce similar affinities in Fc $\gamma$ R SPR binding assays







## LSA<sup>XT</sup> and 8K produce similar on/off rates in trastuzumab Fc $\gamma$ R binding assays

- Kinetic constants within 2-fold across instruments
- CD64 differences likely due to chip chemistry differences

	$k_a$ (1/Ms)	
	Biacore 8K	Carterra LSA <sup>XT</sup>
Hu CD16a V	$(2.6 \pm 0.6) \times 10^5$	$(2.2 \pm 0.1) \times 10^5$
Hu CD16a F	$(1.1 \pm 0.0) \times 10^5$	$(1.2 \pm 0.2) \times 10^5$
Cy CD16	$(2.1 \pm 0.4) \times 10^5$	$(1.2 \pm 0.1) \times 10^5$
Hu CD64	$(7.7 \pm 0.1) \times 10^4$	$(2.1 \pm 0.2) \times 10^5$
Cy CD64	$(1.1 \pm 0.0) \times 10^5$	$(1.4 \pm 0.1) \times 10^5$

	$k_d$ (1/s)	
	Biacore 8K	Carterra LSA <sup>XT</sup>
Hu CD16a V	$(8.4 \pm 1.1) \times 10^{-3}$	$(9.4 \pm 0.3) \times 10^{-3}$
Hu CD16a F	$(2.8 \pm 0.0) \times 10^{-2}$	$(4.4 \pm 0.6) \times 10^{-2}$
Cy CD16	$(1.2 \pm 0.2) \times 10^{-2}$	$(1.2 \pm 0.1) \times 10^{-2}$
Hu CD64	$(1.5 \pm 0.1) \times 10^{-4}$	$(1.8 \pm 0.3) \times 10^{-4}$
Cy CD64	$(7.6 \pm 0.5) \times 10^{-5}$	$(4.5 \pm 0.6) \times 10^{-5}$



## LSA<sup>XT</sup> provides FcγR characterization with greater efficiency than 8K

	<b>Biacore 8K</b>	<b>Carterra LSA<sup>XT</sup></b>
<b>Chip</b>	6 total chips - \$3924	1 chip - \$1190
<b>Analyte</b>	15.9 mg	2.1 mg
<b># of experiments</b>	6 separate experiments	2 combined experiments
<b>Analyst Time</b>	9 hours	3 hours
<b>Replicates</b>	4	32
<b>Cost/replicate</b>	\$89.18	\$3.10
<b>Analyte/replicate</b>	361.4 μg	5.5 μg



## Conclusion

- Carterra LSA<sup>XT</sup> can provide highly accurate and precise Fc $\gamma$ R data using less overall analyte than Biacore 8K.
- Similar kinetic rate constants and K<sub>D</sub> values were recorded between instruments.
- LSA<sup>XT</sup> can perform Fc $\gamma$ R assay faster and cheaper:
  1. Requires 7.5x less analyte
  2. Saves 6+ hours of analyst time
  3. Saves ~\$3,000 in chip costs
  4. Produces 8x as many replicates



## Acknowledgements

### Carterra

- Noah Ditto – [NDitto@Carterra-Bio.com](mailto:NDitto@Carterra-Bio.com)
- Rebecca Rich – [RRich@Carterra-Bio.com](mailto:RRich@Carterra-Bio.com)

### ACRO Biosystems

- Larry Yu – [Larry.Yu@ACROBiosystems.com](mailto:Larry.Yu@ACROBiosystems.com)

### Dragonfly Therapeutics

- Dan Fallon

STP



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# Reference slides

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## FcyRs produced and provided by ACRO Biosystems

Target		Cat No.	Lot No.	Aliquot size
huCD16a V176	FcyRIIIa V176	CDA-H82E9	BV1899-21CRF1-16N	25 µg
huCD16a F176	FcyRIIIa F176	CDA-H82E8	BV1898-21BRF1-12S	25 µg
cyCD16	FcyRIII	FC6-C82E0	BV2520-79XF1-12P	25 µg
huCD16b NA1	FcyRIIIB	CDB-H82E4	BV2424-215KF1-12V	25 µg
huCD16b NA2	FcyRIIIB	CDB-H82Ea	BV2381-224XF2-163	25 µg
huCD32aH	FcyRIIaH	CDA-H82E6	BV1896-222QF1-13V	25 µg
huCD32aR	FcyRIIaR	CDA-H82E5	BV5439A-21BAF1-14Y	25 µg
huCD32b	FcyRIIB	CDB-H82E0	BV1900-2176F1-15H	25 µg
cyCD32b	FcyRIIB	CDB-C82E4	BV3105-93CF1-Y7	25 µg
huCD64	FcyRI	FCA-H82E8	CBV419P1-207EF1-14Y	25 µg
cyCD64	FcyRI	FCA-C82E8	CBV312P1-217FF1-12V	25 µg



## References

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Forest-Nault, C., Gaudreault, J., Henry, O., Durocher, Y., & De Crescenzo, G. (2021). On the Use of Surface Plasmon Resonance Biosensing to Understand IgG-FcγR Interactions. *International Journal of Molecular Sciences*, 22(12), 6616. <https://doi.org/10.3390/ijms22126616>

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Liu, R., Oldham, R. J., Teal, E., Beers, S. A., & Cragg, M. S. (2020). Fc-Engineering for Modulated Effector Functions—Improving Antibodies for Cancer Treatment. *Antibodies*, 9(4), Article 4. <https://doi.org/10.3390/antib9040064>