#### High-Throughput Nanobody Design: Integrating Machine Learning with Rapid Cell-Free Expression

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# ordaos



# Outline

#### 1. Ordaos Technology & Strategy

- $\rightarrow$  Lab-in-the-loop AI-derived binders against disease-specific targets
- $\rightarrow$  Nanobodies and their Advantages
- $\rightarrow$  Cell-Free Expression (CFE)

#### 2. Rapid Screening

- $\rightarrow$  Carterra LSA® platform:
  - High-throughput screening with correlated crude/pure binding data
  - Epitope binning: Evaluating binding/competition profiles
- $\rightarrow$  Wet Lab QC

#### 3. In Vitro Evaluations

 $\rightarrow$  Cell binding assays: Confirm binding in biologically relevant contexts

#### 4. Engineering Function

→ Strategic multimerization: Bispecifics/multivalents to bridge targets and drive activity

# Ordaos Reimagines Biologics Discovery As Biologics Design

Lab-in-the-loop Genai-designed Mini-proteins & Single Domain Antibodies



## Ordaos provides Nanobody and De Novo Protein Solutions

Ai-driven Small Protein Solutions For Next-generation Biologics



# Nanobody Therapeutics: Key Advantages at a Glance

Biophysical Features Enabling Novel Therapeutic Modalities



**Caplacizumab (Cablivi)** Bivalent (homodimeric) Nanobody

- Genetic simplicity → A single cDNA (≈ 400 bp) encodes the complete binding unit. No need to co-express and correctly pair heavy + light chains.
- Very small size (~15 kDa) → superior tissue penetration (including blood-brain barrier) and the ability to reach smaller epitopes
- High physicochemical stability  $\rightarrow$  withstands extreme pH, temperature,
- freeze-thaws; rapidly refolds, enabling room-temperature storage
- Low-cost, high-yield microbial production  $\rightarrow$  expresses efficiently in E. coli, yeast,
- and CFE (no need for chaperones/expensive mammalian cell culture)
- Modular engineering → simple genetic fusions create multivalent, multi-specific molecules, e.g. bi-specific formats, albumin-binding formats to improve half-life
  Reduced immunogenic risk → humanized VHHs resemble human VH3 domains
- and lack an Fc region, lowering possibility for immune side effects
- Regulatory proof-of-concept → FDA-approved Caplacizumab shows nanobodies can meet safety, efficacy and manufacturing standards

# Constant Scaffold with ML-Driven CDR Diversification

Backbone And Tags Stay Fixed While Machine-learning Guides Sequence Variation Only Within CDR1-3



# Ordaos Proprietary Cell-Free Expression Platform

From -80 °C Freezer Stocks To Functional Protein In < 2 hours



- Rapid Protein Generation  $\rightarrow$  60–90 min expression run; complete freezer-to-protein cycle in under two hours.
- Cost-Efficient & In-House Controlled  $\rightarrow$  reduces per-sample cost and gives greater flexibility to optimize reaction conditions.
- Engineered E. coli Lysate  $\rightarrow$  Mechanically lysed e. coli strain engineered for high soluble yield.

•Minimal Reagent needed  $\rightarrow$  20 µL-scale reactions (ng production) produce enough protein for Carterra screens.

- Flexible DNA template  $\rightarrow$  Accepts both linear DNA or plasmids.
- Handles "Difficult" Targets  $\rightarrow$  Toxic, membrane, or

aggregation-prone proteins expressed without the constraints of live-cell growth.

• Easily Scalable & Automatable  $\rightarrow$  2–50 µL reactions in 96- or 384-well plates; compatible with liquid-handling robots.

#### Cell-Free-Expressed Caplacizumab Binds vWF with Native Affinity

Purified CFE Product Yields Correctly Folded, Active, Disulfide-bonded VHH



Lane 1: Mammalian produced VHH Lane 2: Ladder Lane 3: CFE produced VHH; C-tag Purified  Functional CFE Nanobodies → CFE-produced Caplacizumab (nanobody monomer), binds to von Willebrand factor (vWF), demonstrating the capability of our CFE system to produce properly folded, disulfide-formed, functional nanobodies

#### Our fully integrated RL-guided GenAI tailors proteins to disease specifictargets



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# Optimizing Linker and Tag Strategies for Enhanced SPR Performance and Purification

Maximizing SPR Retention and Purification Efficiency through High-Affinity Tag



Changing type of spacer/linker



Changing order and length of linker

•Linker Optimization  $\rightarrow$  Discovered that the 3xHA tag outperforms the 1xHA tag in terms of SPR performance.

- Achieved picomolar capture affinity for our Nanobodies with 3xHA tag
- Excellent retention on SPR chip, even remains after glycine pH 2.0 regeneration.

Tag Order Doesn't Impact Performance → Modulating tag order had minimal effect on retention and affinity.
 C-Tag for Purification → Used for protein purification, also providing high specificity and affinity with our method. We may pivot this tag for capture as well, as the anti-Ctag VHH is extremely stable/regeneratable

#### **Iterative ML-Driven Binder Optimization**

Iteration Strategy: Computational design, rapid screening, and ML feedback in a closed loop to Refine binders



## Ordaos Utilizes Crude CFE Screening with Carterra LSA

Shorter Iteration Time Means More Data And Faster Improvement



#### High-Quality Binder Screening on Carterra LSA with µL-Scale CFE Lysate

#### High-Throughput Assessment of Candidates is possible with Carterra LSA

![](_page_12_Picture_2.jpeg)

![](_page_12_Picture_3.jpeg)

- Crude binding done with 20 μL CFE material and 1 μL DNA
  - 2 wells per plate used for negative and positive control
- Duplicates performed for all samples enabling 188 candidates tested per run

#### Nanodiscs: GPR Binder Screening on Carterra LSA with µL-Scale CFE Lysate

#### Binder screening against difficult membrane-bound GPCR targets

![](_page_13_Picture_2.jpeg)

![](_page_13_Picture_3.jpeg)

- Nanodisc prep  $\rightarrow$  GPCRs reconstituted in-house or sourced commercially, preserving native conformation for SPR.
- Crude lysate screen  $\rightarrow \mu$ L-scale CFE lysate identifies high-affinity, GPCR-specific binders

#### Ordaos generates validated proteins from AI designs in days

SHORTER ITERATION TIME MEANS MORE DATA AND FASTER IMPROVEMENT

![](_page_14_Figure_2.jpeg)

# Crude Lysate Screening Accurately Predicts Purified Protein Binding

Comparable Kinetics Observed With Both Crude And Purified Formats

![](_page_15_Figure_2.jpeg)

Carterra LSA Crude Lysate vs Purified Binding Affinity

• Strong agreement  $\rightarrow$  (R = 0.96;  $\rho$  = 0.92) between KD measured in CFE crude lysate and the same binders after purification via c-tag

- High correlation means we can rank candidates directly from crude reactions, then purify only the top hits for confirmation
- Validates crude-lysate SPR as a reliable, high-throughput strategy for our ML-driven optimization loop
- Saves time and materials → Cuts days and consumables out of each design cycle while maintaining data quality

#### Epitope Binning to Characterize Nanobody Binding Profiles

Assessing Epitope Overlap and Competition with Natural Ligands and Commercial Antibodies

Natural Ligar latural Li 5 Deriv ORD Lea ORD 5 Deriv2

• Binning  $\rightarrow$  Carterra LSA groups nanobodies into distinct epitope communities based on competition profiles

• We can also understand potential steric clashes between nanobody candidates that compete but are known to have distinct epitopes based on HDX data

 Natural Ligand & Commercial Benchmark→ With epitope binning, we can see whether our nanobodies compete/share epitopes with commercial antibodies as well as the target's natural ligand

• Candidate Selection  $\rightarrow$  Informs rational combinations and bispecific designs based on epitope relationships.

#### Cell Binding Assessment of Purified Nanobody Candidates

Flow Cytometry to Confirm Target Engagement on Cells

![](_page_17_Figure_2.jpeg)

 Validated & Purified Binders → Among binders that bind and pass QC thresholds, candidates advance to cell-binding assays to determine whether binding is also seen *in vitro*

#### Strategic Multimerization to make Functional Molecules

Bispecific Reformatting and Avidity Optimization Yield Clinically Relevant Functional Responses

![](_page_18_Figure_2.jpeg)

# Nanobodies against different protein targets

- Multivalency Enables Function → In cases where bridging of multiple proteins is required for function, we engineer multivalent molecules
  - Nanobodies are selected with kinetics matched to cell surface target density
  - Linker configurations guided by in silico modeling

#### Summary

- ML-Guided Binder Design → Rapid generation of high-affinity candidates against disease targets.
- High-Throughput Cell-Free Expression  $\rightarrow$  Produce and screen crude lysates at  $\mu$ L-scale
- Carterra LSA-Powered Screening → Identify hits with crude-to-pure correlation and map epitope diversity via binning
- $QC \rightarrow$  Evaluate stability, producibility, aggregability of binders.
- In Vitro Binding Validation → Determine cell-binding activity and prioritize candidates with therapeutic potential.
- Functional Engineering → Optimize via multimerization or fusion strategies to enhance avidity and mechanism-specific activity.

# Thank you! Let's Connect!

![](_page_20_Picture_1.jpeg)

![](_page_20_Picture_2.jpeg)

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Human-enabled

Machine-driven De novo designed miniproteins to help drug hunters deliver life-saving treatments.